

*Research Article***Targeting Apoptosis as a Therapeutic Approach in Cancer****Rania A. Abd-Elhamid\***, **Maiiada H. Nazmy\*\*** and **Moustafa Fathy\*\*\*\*\***

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**Abstract**

Cancer is a major health problem worldwide, it has been known in its ability in evading and escaping apoptosis. All types of cancer cells gain resistance to apoptosis; they are prone to escape the normal cellular growth and death pathways result in uncontrollable division and proliferation. Apoptosis is programmed cell death or cell suicide; a process normally found in every eukaryotic cell. Damaged DNA triggers the cell to activate apoptotic machinery to remove itself from the body. Apoptosis is an extremely organized process: the cellular membranes are disrupted, the chromosomes are degraded, the DNA breaks up into fragments, and the dying shrinking cell is engulfed by immune cells in a clean and orderly manner. In this review, we will discuss the apoptotic pathways that can be used as a therapeutic target in cancer.

**Keywords:** apoptosis, apoptotic pathways, cancer, p53, cell cycle, cancer stem cell, miRNA.**Introduction**

For decades, cancer has been one of the primary causes of death in all countries all over the world. According to the WHO, there were around 9.6 million deaths due to cancer in 2018 and 70% of such deaths occurred in low and middle income countries.

Currently, cancer management has multiple treatment strategies available; chemo- and radiotherapy and there is also surgery<sup>(1)</sup>. Each treatment strategy has its significance and effectiveness in tumor cells which can be assessed by the ability to initiate the apoptosis cascade, thus, manipulate and restore the overexpressed anti-apoptotic proteins levels, and increase the expression of pro-apoptotic molecules could be the new goal of treatment. Recognizing the underlying mechanism of programmed cell death has created a new insight in cancer treatment by obtaining specific molecules that aim for a signal, gene and/or protein in cancer cells to promote its suicidal; meanwhile, non-harmful to the normally developed cells. In this article, we evaluate how apoptosis can be used as targeted therapy in cancer.

**Apoptosis**

Programmed cell death; apoptosis is a normal physiological process which is critical for life.

Its significance relies in tumor suppression, maintaining tissue homeostasis and infection resistance. The balance between cell proliferation and death must be maintained throughout life<sup>(2)</sup>. Apoptosis; a strictly controlled complex process at the molecular level regulated by the balance between the pro- and anti-apoptotic proteins has the liability of clearing any damaged DNA containing cells out of the body<sup>(3)</sup>. Both, the extrinsic and intrinsic initiators of the apoptotic pathways which lead to activation of caspases cascade conduct several and complete characteristic changes of cell morphology that undergoing apoptosis<sup>(4)</sup>.

**Morphological and Biochemical Changes in Apoptosis**

Morphologically, apoptotic cell undergoes lessening in size, budding of plasma membrane while maintaining its integrity till the end, nucleic acid degradation and expression of phosphatidylserine on the outer layers of the membrane<sup>(5, 6)</sup> which allows early phagocytosis by macrophages to recognize, detect and engulf apoptotic cells in clear distinction manner than necrosis which usually accompanied by the presence of pro-inflammatory signals that would lead to cellular lysis and inflammation<sup>(7)</sup>.

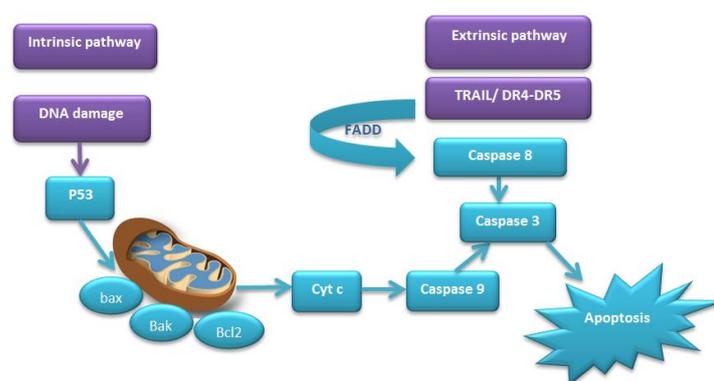
The organelles within the cells are prone to specific alterations in the morphological

features that occur in the late stages of apoptosis, membrane blebbing can also be observed and the change in its asymmetry<sup>(8)</sup>. Caspases; a family of proteases which is the main regulators and key players of apoptosis is synthesized as pro-caspases; inactive proteins<sup>(9)</sup>.

They are classified into initiators and executors/effectors. Initiators such as: caspase-2,-8, and 9 begin the apoptosis mechanism through the auto-activation and auto-cleavage of pro-caspases by an auto-proteolysis process, while executors such as caspase-3, -6 -7 and -10 proteolyze target substrate for the purpose of cell death. Since initiators are responsible for

the beginning of the apoptosis mechanism, they start by binding to specific adaptor molecules to become activated<sup>(10, 11)</sup>.

Then, they activate executors' caspases which result in cytoplasmic endonuclease activation and chromatin condensation. This highly strict irreversible step represent the optimum goal of both extrinsic and intrinsic pathways<sup>(12)</sup>. This profound mechanism controlled by caspases is responsible of the features and morphological changes that occur during apoptosis, such as plasma budding, cytoplasmic blebbing and apoptotic bodies<sup>(13)</sup>. **(Figure 1)**



**Figure (1): Graphical illustration of intrinsic and extrinsic apoptotic pathways.**

### Targeting Bcl-2 Family

One of the foremost vital pathways of apoptosis is mitochondria-dependent intrinsic pathway. Thus, targeting this pathway is practical approach to induce apoptosis<sup>(14)</sup>. Bcl-2 family is the key player of the intrinsic pathways, which either promote or suppress the permeability of mitochondrial membrane as needed for the release or blocking of cytochrome-c and other apoptotic proteins<sup>(15, 16)</sup>.

Since anti-apoptotic proteins work by inhibiting apoptosis and increasing the viability and the proliferation of the cells, it was reported in several studies that their over-expression is in correlation with tumor progression and development<sup>(17)</sup>.

Bcl-2 antisense (oblimersen sodium) is the first drug reported to target Bcl-2 pathway that entered clinical trials, Bcl-2 antisense decreased the mortality and increased the survival of

chronic lymphocytic leukemia patients by increasing the sensitivity to chemotherapy drugs when both used in combination<sup>(18, 19)</sup>.

Another study suggested the use of ABT-263 (navitoclax) to target bcl-2 family which showed efficacy in clinical trials. It was suggested that combination strategy between ABT-263 with anthracyclines such as doxorubicin or with cyclin dependant kinase 9 inhibitors such as dinaciclib induced potent apoptosis in small cell lung cancer cell lines<sup>(20)</sup>. The use of navitoclax is limited by its major side effects of thrombocytopenia<sup>(21)</sup>.

BH3 mimetic anti-cancer drug has been approved by the FDA in 2016 under the name of venetoclax for treating chronic lymphocytic leukemia. BH3 mimetics bind to the bcl-2 freeing BIM. The freed BIM can then bind and sequester any unoccupied non-targeted anti-apoptotic protein and facilitates apoptosis<sup>(22)</sup>.

This drug does not cause thrombocytopenia while retaining proapoptotic activity<sup>(21)</sup>.

Navitoclax and venetoclax have broad specificity against bcl-2 but not MCL1 (bcl-2 family member) as it has significant different binding site than bcl2. Obtaining a drug would target MCL1 is a target as it is one of the most commonly amplified genes across all cancer types<sup>(23)</sup>.

The AM-8621 and VU661013, a spiromacrocyclic ring with submolar binding affinity to the MCL1 groove displace BIM from MCL1, with lower binding affinity to BCL2 or BCL-xL<sup>(24)</sup>. The disturbing in the interaction between BIM-MCL1 lead to activation of BAK and caspases and cause apoptosis induction.

Ramsey and his colleges showed the beneficial effect of using BCL2 and MCL1 inhibitors in sequence or in combination in acute myeloblastic leukemia clinical trials to induce apoptosis<sup>(25)</sup>.

Also, small-molecule tyrosine kinase inhibitors (erlotinib, lapatinib) used to increase the sensitivity of navitoclax-mediated apoptosis, which caused MCL1 degradation. This increased MCL1 degradation can induce dramatic apoptotic responses<sup>(26)</sup>.

Nangia and his colleges indicated that the combination therapy between MEK and MCL1 inhibition induces apoptosis and tumor suppression in KRAS mutant non-small cell lung cancer (NSCLC) which was synergized by the exposure to BCL-XL inhibitors at first, which promotes the binding of pro-apoptotic BCL-2 proteins to MCL-1<sup>(27)</sup>.

Graviola is a plant derived compound has been shown anticancer properties. A study revealed it inhibit BCL-2 proteins while increasing BAX and promoting apoptosis<sup>(28)</sup>. The exact mechanism still not completely understood, but it shows promising activity as potential treatment in cancer.

### Targeting the Tumor Suppressor Gene P53

There are multiple anticancer drugs would target p53 mediated pathways as it is an important tumor inhibitor and also proapoptotic protein that would initiate apoptosis. For decades, p53 has been known for its ability

to initiate DNA repair by increasing the expression of specific target genes through binding to their regulatory sequences<sup>(29)</sup>.

It is primary goal to repair damaged DNA, but if DNA is extensively damaged, cell cycle arrest or apoptosis will occur due to the activation of p53<sup>(30)</sup>. P53 function is essential to inhibit tumorigenesis.

Thus, many new anti-cancer strategies target the stimulation or the refolding of the mutant p53 pathway to induce cell death<sup>(31)</sup>. This could be by targeting p53 itself or by using small molecules capable of restoring p53 functions. Also, aiming its target gene products, such as the pro-apoptotic Bax protein, which show ability to mediate apoptosis, inhibit Bcl-2 expression and initiate the intrinsic apoptotic pathway<sup>(32, 33)</sup>.

P53 mutation and inactivation is observed in different tumor tissue, cells biopsies and cell lines which represent a relation between p53 mutation and tumor progression<sup>(34)</sup>.

Based on a recent study by Sergiu and co-workers investigated the use of CRISPER/Cas system to restore the TP53 in tumor cells by completely replacing the mut-p53 with the functional copy in the human genome showed promising results in induction of cell suicide and apoptosis<sup>(35)</sup>.

A compound that represents a breakthrough in the activation of mutant p53 thiosemicarbazones which was investigated by Yu and his colleges served as source of zinc for mut-p53. Wt-p53 contains zinc ion which is essential for the binding between the p53 and DNA domain. This drug allowed refolding of the mutant p53 to its wild type conformation and caused proper binding between the p53 and the DNA<sup>(36)</sup>.

Also, testing polyarginine on p53 mutant cancer cells in vitro has been investigated by Kana-pathipillai. The results showed significant inhibition of polyarginine and polyornithine on p53 mutant peptide aggregation in vitro and growth inhibition of p53 mutant (R248Q) lung cancer cells H719 and p53 mutant (R175H) breast cancer cells SK-BR-3, with no effect on the wild type p53 and exhibit no toxicity on normal cells<sup>(37)</sup>.

According to Wiman and co-workers, the use of PRIMA-1 (p53-Reactivation and Induction of Massive Apoptosis-1); quinuclidine compound initiated the DNA binding activity in vitro in various cancer cells bearing mut-p53. This drug increased the expression of p53 target genes such as BAX, PUMA and NOXA and also activated caspases for apoptosis induction, and its methylated derivative; PRIMA-1Met caused better and promising results<sup>(36)</sup>.

Another drug for restoring the function of mut-p53 is MIRA-1 (mut-p53-dependent induction of rapid apoptosis). Although the chemical structures of MIRA and PRIMA-1 are distinct from each other, they have quite similar molecular mechanism for the re-activation of mut-p53<sup>(38)</sup>.

They both alter thiol groups, while MIRA-1 induces cell death, caspase activation and apoptosis in mut-p53 dependant pathway much faster with higher potency than PRIMA-1<sup>(39)</sup>.

According to a study by Sidharta Chatterjee, the scientist designed bioactive compound to block the signaling pathway between HDM2/MDM2 and p53 to increase p53 expression in cancer cells for apoptosis induction. MDM2 is a negative regulator for p53: it induces p53 fragmentation and inhibits p53 activity as a result of a binding between MDM2 and p53<sup>(40)</sup>.

### Targeting the Extrinsic Cell Death

Pro-apoptotic membrane receptors; death domain (DD)-containing receptors present on the cell surface such as DR4 and DR5 binds to specific pro-apoptotic ligands such as TNF related apoptosis-inducing ligand (Apo2L/TRAIL) in order to initiate and activate the extrinsic apoptotic pathway<sup>(41)</sup>.

A series of cascades is activated once the receptor is bound to a specific ligand; it starts activation of the death-inducing signaling complex (DISC) through utilizing the adapter Fas-associated death domain (FADD) and procaspases 8 and 10, which result in activation of caspase cascades<sup>(42)</sup>.

The Apo2L/TRAIL pathway is a promising approach in cancer therapy because of its integrity in different types of cancer, induction of apoptosis in cancer cells with no effect on normal cells and irrelative to p53 function<sup>(43)</sup>.

Utilizing the natural ligands that selectively activate pro-apoptotic receptors such as DR4 and DR5 has been applied in the market. The synthesis of soluble recombinant human protein Apo2L/TRAIL was developed, moreover; Genentech has developed various monoclonal antibodies that also act on pro-apoptotic receptors such as the fully human DR5 agonistic antibody Apomab<sup>(44)</sup>.

Recombinant human TRAIL (rhTRAIL); which known as dulanermin is TRAIL receptor agonist that showed promising activity in apoptosis induction in cancer cells<sup>(45)</sup>.

rhTRAIL is a soluble small protein binds to DRs with broader activity to maintain cytotoxicity but also with lack of specificity. There are numerous rhTRAIL were developed, for example: leucineoriso-leucine zipper TRAIL (lz-TRAIL, iz-TRAIL), hexahistidine-TRAIL (6xHis-TRAIL), FLAG-TRAIL, and Tenascin-C TRAIL (TNC-TRAIL). They demonstrated higher stability than dulanermin but showed toxicity on normal tissue and cells<sup>(46)</sup>.

Another approach was obtained is the fusion of trimeric TRAIL chain to a human IgG1 crystallizable fragment (Fc) or to single-chain variable antibody fragment (scFv) which resulted in Fc-TRAILs and sc-TRAILs, respectively. This approach showed better results in apoptosis induction and stability than dulanermin<sup>(47)</sup>.

Wang et al., investigated the cytotoxic activity of small molecule known as biomifi or (Z)-5-(5-[(3-[4-bromophenyl]-2-imino-4-oxothiazolidin-5-ylidene) methyl] furan-2yl) isoindoline-1,3-dione<sup>(1)</sup>.

Biomifi has good binding affinity to DR5 with little affinity to DR4, it showed potential cytotoxic activity on lung cancer cells H460 and H1155, the osteosarcomacellline U2OS, glioblastoma cells T98G, cervical cancer cells HeLa and colon cancer cells HT29<sup>(48)</sup>.

Allen and co-workers introduced TRAIL inducing compound 10 (TIC-10); a drug was found to inhibit ERK and PI3K/Akt pathways and up regulate TRAIL receptors for apoptosis induction. This drug was also found to cross

blood brain barrier with great potential to treat central nervous system tumors<sup>(49)</sup>.

Various other analogues were developed such as a trifluoromethylbenzyl congener and a difluorobenzyl analogue with greater cytotoxic activity and minimal/no effect on normal cells<sup>(50)</sup>.

The use of ibulocydine; a synthetic drug and TRAIL mediated cell death. It is cyclin-dependant kinas inhibitor that suppresses Cdk7 and Cdk9 on cancer cells to induce apoptosis through activating caspases<sup>(51)</sup>.

Another study showed that 2-deoxy-D-glucose induced apoptosis through TRAIL mediated pathways in human gastric tumor cells<sup>(52)</sup>.

Numerous of drugs have been used before to induce the apoptotic process of DRs and ligands in the extrinsic pathway, such as cyclooxygenase-2 inhibitors, histone deacetylase (HDAC) inhibitors, proteasome inhibitors and a number of antibodies which target the DR<sup>(53)</sup>.

Various approaches proposed the cFLIP (cellular FLICE-like) inhibitory protein as a therapeutic target in cancer for apoptosis induction. Some studies used siRNA to inhibit the anti-apoptotic effect of cFLIP and increase the sensitivity to TRAIL-mediated pathway and/or other chemotherapeutic agents. However; these approaches showed some limitations starts from the safe delivery of the siRNA and significant homology between cFLIP and caspase-8<sup>(54)</sup>.

Thus, TRAIL mediated apoptosis represent an effective approach in cancer therapy, the pre-clinical and clinical trials showed safety and significant effect for apoptosis induction.

### Targeting Convergence Pathway

A merge occur between both extrinsic and intrinsic pathways of apoptosis on downstream series of caspases, it is named the execution phase which is the final pathway of apoptosis. It has been indicated that apoptosis could be suppressed at this step.

The apoptosis-inhibiting proteins (IAPs) is an evolutionarily conserved family of apoptosis

suppressors, which has been shown to influence cell death<sup>(55)</sup>.

Poor prognosis of cancer was reported with over-expression of IAPs<sup>(56,57)</sup>. All IAPs family can bind and inhibit caspases function which allows them to inhibit apoptosis through the so-called BIR (baculovirus iap repeat) domain, which facilitate protein-protein interactions and prevent the conversion of inactive pro-caspases to active caspases<sup>(58)</sup>.

Caspase-3 & 8 couldn't be processed and activated in response to IAPs, thereby extrinsic apoptotic signaling was inhibited<sup>(59)</sup>.

Several studies extensively indicated the use of IAPs in anticancer therapeutics and proved its efficiency in targeting and initiating cell suicide<sup>(60)</sup>.

Since their mechanism depend mainly in a balance between IAPs and their endogenous antagonists which keep them in order and promote apoptosis. Thus, the use of naturally occurring IAP-antagonists such as SMAC (Diablo) and HtrA2 (Omi) to aim and bind to IAPs in order to pave the way for caspases to initiate apoptosis. These naturally occurring IAP-antagonist are segregated in the mitochondria and liberated to the cytoplasm during apoptosis<sup>(61)</sup>.

There is some cytotoxic anticancer drugs use synthetic peptidesbased that imitate the action of SMAC and HtrA2 to induce cell death or sensitize cancer cells to apoptosis<sup>(62)</sup>. Other IAP protein inhibitors have been developed such as: SH122, SH130, SM164, AZD5582, JP1201, AEG35156, LY2181308 and YM155<sup>(63)</sup>.

### Initiating Cell Cycle Arrest to Induce Apoptosis

One of the cancer drug target approaches has been investigated is the cyclin-dependent kinases (CDKs) family that control and monitor the cell cycle phases. For decades, CDKs was known as the main contributors of cell cycle, development, proliferation and differentiation.

However; recently, potential targets such as protein kinases that act upon DNA damage and

protein kinases that govern mitosis were identified<sup>(64, 65)</sup>.

Cell cycle arrest occurs as a result of DNA lesions give the cell a chance to repair damaged DNA. The arrest of cell cycle obtained by two essential checkpoints- 1 and 2 through the activation of serine– threonine protein kinase are anticipated to maintain DNA integrity and act as self-cell defenses against cancer.

There have been trials for the synthesis of kinases inhibitors as anticancer therapeutics that leads to the synthesis of BCR–ABL protein kinase inhibitor imatinib from Gleevec; Novartis, it successfully target tyrosine kinases for the treatment of chronic myelogenous leukaemia. Several studies approved that aiming CDK family members is a good addition to the list of therapeutics- inducing apoptosis, particularly for CDK-2, 4 and 5. Other study suggested that CDK1 is considered a potential approach in prostate cancer, while CDK5 has a particular role in regulating cell motility and metastatic in such disease<sup>(66)</sup>.

Moreover, a good understanding about the CDK family, particularly the relation between their regulators and their inhibitors have paved the ways for new strategies' targeting apoptosis<sup>(64)</sup>.

A study recently done by Imran Khan and colleges investigated the effect of carvacol; a flavonoid found abundantly in thyme plants. They found that carvacol efficiently cell cycle arrest by suppressing CDK-2,-4,-6, cyclin E and cyclin D1 and increasing p21 expression<sup>(67)</sup>.

Zhang et al., proposed the combined use of CDK1 inhibitor RO-3306 or dinaciclib with cobimetinib (MEK inhibitor) caused cell cycle arrest and apoptosis in human colorectal cancer<sup>(68)</sup>.

Humeau and co-workers investigated the role of Ca<sup>2+</sup> in cell cycle and cell proliferation and suggested targeting Ca<sup>2+</sup> as an approach for cell cycle arrest and apoptosis induction<sup>(69)</sup>. Ca<sup>2+</sup> is crucial for numerous physiological processes include cell differentiation, proliferation and cell death and cytoplasmic Ca<sup>2+</sup> levels varies along the cell cycle. Drugs targeting Ca<sup>2+</sup> such as Ca<sup>2+</sup> channel blockers can

be used to target Ca<sup>2+</sup> channels that is over expressed in cancer, for example: T-type Ca<sup>2+</sup> channels, SOCE channel components (ORAI1 and STIM1), InsP3R and RyR channels, also TRPM8, TRPV6 and TRPC1/C4 channels in the TRP family<sup>(70)</sup>.

### Targeting Cancer Stem Cell (CSC) to Induce Apoptosis

Currently, there is evidence that some cancer cells have stem cell like features such as self-renewal, infinite proliferation and replication and the ability to survive toxic agents which considered is the main reason of cancer recurrence after radiation and chemotherapy<sup>(71-73)</sup> thus, the name of cancer stem cell (CSC).

Other properties of CSC include: genetic and chromosomal instability, mobilization of cellular resources, chromatin transcription, epigenetic modifications and altered environmental interactions between cancer cells and normal cells within the extracellular and endothelium<sup>(74)</sup>. CSC can be originated from the transformation of normal stem cells through several genetic mutation and instability, or they can occur from tumor cells acquire stem cells like features progressively<sup>(75, 76)</sup>.

Targeting and eradicating CSCs through apoptosis is a therapeutic goal for cancer and holds a promising approach to decrease morbidity and mortality in cancer patients. Therefore, many compounds have been developed to target intrinsic and extrinsic apoptotic pathways.

A combination between TRAIL and cisplatin was reported to eradicate CSCs effectively<sup>(77)</sup>. For example, it showed great potentials in enhancing triple negative breast cancer stem cells death and apoptosis through suppression of Wnt signaling pathway<sup>(78)</sup>. It was also reported that the co-treatment between TRAIL and cytarabine or daunorubicin inhibited the growth of progenitor cells in acute myeloid disease<sup>(79)</sup>.

Moreover, when TRAIL is used in addition to Bortezomib, a proteasome inhibitor, it initiated apoptosis in glioblastoma stem cells<sup>(80)</sup>. Furthermore, a study showed that injecting mesenchymal stem cells (MSC) engineered to express TRAIL into mice resulted in inhibition

of tumor growth and induction of apoptosis in squamous and lung cancer stem cell<sup>(81)</sup>.

Another pathway might be initiated for the purpose of CSCs death and apoptosis is NF- $\kappa$ B, a transcription factor with a critical role in apoptosis signaling pathway. In fact, NF- $\kappa$ B suppress the programmed cell death and promotes cell proliferation, tumorigenesis and metastasis<sup>(82)</sup>.

Small molecules have been developed to inhibit NF- $\kappa$ B like parthenolide, pyrrolidinedithiocarbamate and its analog diethyldithiocarbamate. They showed a promising result in targeting breast cancer stem cells which indicate the vital activity of NF- $\kappa$ B to promote the proliferation cancer stem cells<sup>(83)</sup>.

Other strategy occurred through the co-treatment between the proteasome inhibitor MG-132 together with the anticancer drug idarubicin induced apoptosis in leukemic stem cells through the suppression of NF- $\kappa$ B<sup>(84)</sup>.

Recent in vivo and in vitro studies suggested the use of dietary phytochemicals to interfere in signaling transduction pathways<sup>(85-87)</sup>, thus, to induce apoptosis and inhibit proliferation and cell cycle progression in tumor cells and particularly in CSCs.

Curcumin was shown to regulate miRNA expression in breast cancer stem cells, it inhibited Bcl-2 expression with the induction of apoptosis, it showed no effect on normal stem cells<sup>(88, 89)</sup>, curcumin in combination with resveratrol (substance found in grapes) also showed inhibition in Wnt signaling and promising targets of the self-renewal features of CSCs<sup>(90)</sup>.

Isothiocyanates was reported as potent anti-cancer phytochemical. Their role depends on induction of apoptosis of CSCs and cell cycle arrest<sup>(91)</sup>.

Sulforaphane; a phytochemical found in broccoli was reported to target pancreatic CSCs through NF- $\kappa$ B<sup>(92)</sup>, it was also reported to target breast cancer progenitor stem cell through Wnt/ $\beta$ -catenin self-renewal pathway<sup>(93)</sup>.  $\beta$ -Carotene has been recognized to suppress the growth of CSCs in neuroblastoma<sup>(94)</sup>.

A recent study by Jian-feng Li and his colleagues targeted lung cancer stem cells by the use of bispecific antibody (BsAbs) which were able to block two different antigen c-MET and CTLA-4<sup>(95)</sup>.

Their theory based upon the up regulation of Cellular mesenchymal-to-epithelial transition factor (c-MET) and cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) expression contributes to promoting tumorigenesis in different types of solid tumors, and targeting both antigens should suppress tumor progression through the inhibition of hepatocyte growth factor (HGF) mediated tumor development<sup>(96)</sup>. HGF is cytokine present in a variety of with anti-apoptotic activities<sup>(97)</sup>, thus, its inhibition would promote CSCs apoptosis.

Deregulation of PI3K/AKT/mTOR signaling is found in many cancers and the one of the reasons of treatment resistance which has a critical role in CSCs<sup>(98)</sup>. The co-treatment of metformin, an anti-diabetic drug with an inhibitor of PI3K/AKT/mTOR signaling, and the RAF inhibitor sorafenib effectively reduce GSC oxidative stress and efflux pump activity, and synergistically CSCs<sup>(99)</sup>. Moreover, a recent report showed that BFZ-235, an inhibitor of PI3K/AKT/mTOR signaling, effectively suppresses the stemness of colon CSCs<sup>(100)</sup>.

Chao shang and his colleges suggested the role of miRNA-21 inhibition to induce glioblastoma cancer stem cells death and apoptosis through targeting FASLG as a new potential therapeutic approach<sup>(101)</sup>.

### Targeting miRNA to Induce Apoptosis

There is an increase attention to microRNAs (miRNAs) recently<sup>(102)</sup>. They are non-coding short RNAs of 18–25 nucleotides in length. They were found to govern gene expression at post-transcriptional level, either by degradation of mRNA or stop the translation process<sup>(103)</sup>. miRNAs contribute in multiple cellular functions, such as cell cycle regulation, cell differentiation, stem cell self-renewal and apoptosis.

Furthermore, it was reported that the decreased expression of miRNA causes eradication of apoptosis, which lead to carcinogenesis and resistance to the therapy<sup>(105)</sup>. Therefore, restoring miRNA levels especially those

responsible for apoptosis signaling could be a promising approach for developing effective treatment against tumorigenesis.

This has attracted the interest of new researchers to intensively develop new drugs to restore miRNA levels. miRNAs can act as oncogenes by which malignantly transform cells into cancer, or tumor suppressor genes with the ability to inhibit tumorigenesis.

It has been reported that miRNAs could be considered in cancer therapeutic strategies. Moreover, it has been intensively studied the potential effect of targeting tumor-inducing miRNAs and restoring the levels of tumor suppressive miRNAs for therapeutic purposes.

There is a strategy of using miRNA antagonists through the virus delivery system of miRNAs with the beneficial effect of the virus not being incorporated into the genetic material. This strategy has shown the beverage of being effective and non-mutagenic.

Another strategy is to introduce miRNA mimic; synthetic double stranded RNA aimed to imitate the endogenous miRNAs as tumour-suppressor genes has been found to induce apoptotic pathways.

Several studies documented the beneficial effect of transfection tumor suppressive miRNAs such as anti-miR-24 oligonucleotides<sup>(112)</sup> to inhibit proliferation and initiate cell suicide<sup>(113)</sup>.

As well, It was also documented that miR-24-2, miR 365-2 and miR-195 affect negatively on the oncogenic anti-apoptotic Bcl2<sup>(114)</sup>. Also, decreased levels of miR-15, miR-16, and let-7 resulted in anti-apoptotic genes activation in different types of cancer cells<sup>(115-117)</sup>.

### Conclusion

The vast majority of what we think about apoptosis has been established in the most recent decade. The idea to objectively target apoptosis signal transduction pathways has significant effect for malignancy treatment, since targeting cell death is essential for the effectiveness and potency of most anticancer treatments. Induction of apoptosis not just legitimately triggers cell suicide, yet in addition increases the sensitivity of tumor cells for apoptosis.

The main goal of therapeutic approaches for malignant growth depends on the way that body can maintain a healthy number of cells, yet, this concept is enormously exasperates in malignancy cells. Distinguishing proof of the key players associated with the apoptosis process and their cooperation with other members of apoptosis has supported the critical improvements made in the field of malignancy treatment.

Cancer therapy based upon initiation and induction of apoptosis has been a key methodology in battling this disease; nonetheless, we are still left with immense difficulties to be defeated.

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### References

1. Tekade RK, Dutta T, Tyagi A, Bharti AC, Das BC, Jain NK. Surface-engineered dendrimers for dual drug delivery: a receptor up-regulation and enhanced cancer targeting strategy. *Journal of drug targeting*. 2008;16(10):758-72.
2. Czabotar PE, Lessene G, Strasser A, Adams JM. Control of apoptosis by the BCL-2 protein family: implications for physiology and therapy. *Nature reviews Molecular cell biology*. 2014;15(1):49-63.
3. Fuchs Y, Steller H. Programmed cell death in animal development and disease. *Cell*. 2011;147(4):742-58.
4. Wong RS. Apoptosis in cancer: from pathogenesis to treatment. *Journal of experimental & clinical cancer research* : CR. 2011;30:87.
5. Hacker G. The morphology of apoptosis. *Cell and tissue research*. 2000;301(1):5-17.
6. Saraste A, Pulkki K. Morphologic and biochemical hallmarks of apoptosis. *Cardiovascular research*. 2000;45(3):528-37.
7. Hengartner MO. The biochemistry of apoptosis. *Nature*. 2000;407(6805):770-6.

8. Kroemer G, El-Deiry WS, Golstein P, Peter ME, Vaux D, Vandenabeele P, et al., Classification of cell death: recommendations of the Nomenclature Committee on Cell Death. *Cell death and differentiation*. 2005;12 Suppl 2:1463-7.
9. Li J, Yuan J. Caspases in apoptosis and beyond. *Oncogene*. 2008;27(48):6194-206.
10. Nicholson DW. Caspase structure, proteolytic substrates, and function during apoptotic cell death. *Cell death and differentiation*. 1999;6(11):1028-42.
11. Fathy M, Fawzy MA, Hintzsche H, Nikaido T, Dandekar T, Othman EM. Eugenol Exerts Apoptotic Effect and Modulates the Sensitivity of HeLa Cells to Cisplatin and Radiation. *Molecules*. 2019;24(21).
12. Stennicke HR, Salvesen GS. Caspase assays. *Methods in enzymology*. 2000;322: 91-100.
13. Degtrev A, Boyce M, Yuan J. A decade of caspases. *Oncogene*. 2003;22(53):8543-67.
14. Hassan M, Watari H, AbuAlmaaty A, Ohba Y, Sakuragi N. Apoptosis and Molecular Targeting Therapy in Cancer. *BioMed research international*. 2014; 2014:23.
15. Green DR, Kroemer G. The pathophysiology of mitochondrial cell death. *Science*. 2004;305(5684):626-9.
16. Kroemer G, Reed JC. Mitochondrial control of cell death. *Nature medicine*. 2000;6(5):513-9.
17. Gautschi O, Tschopp S, Olie RA, Leech SH, Simoes-Wust AP, Ziegler A, et al., Activity of a novel bcl-2/bcl-xL-bispecific antisense oligonucleotide against tumors of diverse histologic origins. *Journal of the National Cancer Institute*. 2001;93(6):463-71.
18. Muchmore SW, Sattler M, Liang H, Meadows RP, Harlan JE, Yoon HS, et al., X-ray and NMR structure of human Bcl-xL, an inhibitor of programmed cell death. *Nature*. 1996;381(6580):335-41.
19. Rai KR. The natural history of CLL and new prognostic markers. *Clinical advances in hematology & oncology: H&O*. 2008; 6(5):4-5; quiz 10-2.
20. Inoue-Yamauchi A, Jeng PS, Kim K, Chen H-C, Han S, Ganesan YT, et al., Targeting the differential addiction to anti-apoptotic BCL-2 family for cancer therapy. *Nature Communications*. 2017;8(1):16078.
21. Hata AN, Engelman JA, Faber AC. The BCL2 Family: Key Mediators of the Apoptotic Response to Targeted Anti-cancer Therapeutics. *Cancer Discovery*. 2015;5(5):475-87.
22. Adams JM, Cory S. The BCL-2 arbiters of apoptosis and their growing role as cancer targets. *Cell Death & Differentiation*. 2018; 25(1):27-36.
23. Beroukhim R, Mermel CH, Porter D, Wei G, Raychaudhuri S, Donovan J, et al., The landscape of somatic copy-number alteration across human cancers. *Nature*. 2010; 463(7283):899-905.
24. Leber B, Kale J, Andrews DW. Unleashing Blocked Apoptosis in Cancer Cells: New MCL1 Inhibitors Find Their Groove. *Cancer Discovery*. 2018;8(12):1511-4.
25. Ramsey HE, Fischer MA, Lee T, Gorska AE, Arrate MP, Fuller L, et al., A Novel MCL1 Inhibitor Combined with Venetoclax Rescues Venetoclax-Resistant Acute Myelogenous Leukemia. *Cancer Discovery*. 2018;8(12):1566-81.
26. Arai S, Jonas O, Whitman MA, Corey E, Balk SP, Chen S. Tyrosine Kinase Inhibitors Increase MCL1 Degradation and in Combination with BCLXL/BCL2 Inhibitors Drive Prostate Cancer Apoptosis. *Clinical Cancer Research*. 2018;24 (21):5458-70.
27. Nangia V, Siddiqui FM, Caenepeel S, Timonina D, Bilton SJ, Phan N, et al., Exploiting MCL1 Dependency with Combination MEK + MCL1 Inhibitors Leads to Induction of Apoptosis and Tumor Regression in KRAS-Mutant Non-Small Cell Lung Cancer. *Cancer Discovery*. 2018;8(12):1598-613.
28. Pfeffer CM, Singh AT. Apoptosis: a target for anticancer therapy. *International journal of molecular sciences*. 2018;19 (2):448.
29. Vogelstein B, Lane D, Levine AJ. Surfing the p53 network. *Nature*. 2000;408 (6810): 307-10.
30. Vermeulen K, Berneman ZN, Van Bockstaele DR. Cell cycle and apoptosis. *Cell Prolif*. 2003;36(3):165-75.
31. Blaydes JP, Craig AL, Wallace M, Ball HM, Traynor NJ, Gibbs NK, et al., Synergistic activation of p53-dependent

- transcription by two cooperating damage recognition pathways. *Oncogene*. 2000; 19(34):3829-39.
32. Elmore S. Apoptosis: a review of programmed cell death. *Toxicol Pathol*. 2007; 35(4):495-516.
  33. Beberok A, Wrzesniok D, Rok J, Rzepka Z, Respondek M, Buszman E. Ciprofloxacin triggers the apoptosis of human triple-negative breast cancer MDA-MB-231 cells via the p53/Bax/Bcl-2 signaling pathway. *International journal of oncology*. 2018;8(10).
  34. Silva JL, De Moura Gallo CV, Costa DC, Rangel LP. Prion-like aggregation of mutant p53 in cancer. *Trends in biochemical sciences*. 2014; 39(6):260-7.
  35. Chira S, Gulei D, Hajitou A, Berindan-Neagoe I. Restoring the p53 'Guardian' Phenotype in p53-Deficient Tumor Cells with CRISPR/Cas9. *Trends in Biotechnology*. 2018;36(7):653-60.
  36. Hientz K, Mohr A, Bhakta-Guha D, Efferth T. The role of p53 in cancer drug resistance and targeted chemotherapy. *Oncotarget*. 2017;8(5):8921-46.
  37. Kanapathipillai M. Treating p53 Mutant Aggregation-Associated Cancer. *Cancers*. 2018;10(6).
  38. Katzenellenbogen BS, Choi I, Delage-Mourroux R, Ediger TR, Martini PG, Montano M, et al., Molecular mechanisms of estrogen action: selective ligands and receptor pharmacology. *The Journal of steroid biochemistry and molecular biology*. 2000;74(5):279-85.
  39. Berger C, Qian Y, Chen X. The p53-estrogen receptor loop in cancer. *Current molecular medicine*. 2013;13(8):1229-40.
  40. Chatterjee S. Design of Novel Putative Peptide fragments derived from exons of p53 isoform  $\alpha$  and p53 promoter region ORF sequence targeting HDM2/MDM2-p53 interaction. 2018.
  41. Ashkenazi A, Dixit VM. Death receptors: signaling and modulation. *Science*. 1998; 281(5381):1305-8.
  42. Locksley RM, Killeen N, Lenardo MJ. The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell*. 2001; 104(4):487-501.
  43. Ashkenazi A, Pai RC, Fong S, Leung S, Lawrence DA, Marsters SA, et al., Safety and antitumor activity of recombinant soluble Apo2 ligand. *The Journal of clinical investigation*. 1999;104(2):155-62.
  44. Fesik SW. Promoting apoptosis as a strategy for cancer drug discovery. *Nature reviews Cancer*. 2005; 5(11):876-85.
  45. Holland PM. Targeting Apo2L/TRAIL receptors by soluble Apo2L/TRAIL. *Cancer letters*. 2013; 332(2):156-62.
  46. Ion GND, Nitulescu GM, Popescu CI. Targeting TRAIL. *Bioorganic & medicinal chemistry letters*. 2019;29(18):2527-34.
  47. Legler K, Hauser C, Egberts JH, Willms A, Heneweer C, Boretius S, et al., The novel TRAIL-receptor agonist APG350 exerts superior therapeutic activity in pancreatic cancer cells. *Cell death & disease*. 2018;9(5):445.
  48. Wang G, Wang X, Yu H, Wei S, Williams N, Holmes DL, et al., Small-molecule activation of the TRAIL receptor DR5 in human cancer cells. *Nature chemical biology*. 2013;9(2):84-9.
  49. Allen JE, Kline CL, Prabhu VV, Wagner J, Ishizawa J, Madhukar N, et al., Discovery and clinical introduction of first-in-class imipridone ONC201. *Oncotarget*. 2016;7(45):74380-92.
  50. Wagner J, Kline CL, Ralff MD, Lev A, Lulla A, Zhou L, et al. Preclinical evaluation of the imipridone family, analogs of clinical stage anti-cancer small molecule ONC201, reveals potent anti-cancer effects of ONC212. *Cell cycle*. 2017; 16(19):1790-9.
  51. Park SS, Jwa E, Shin SH, Ju EJ, Park I, Pak JH, et al., Ibulocydine sensitizes human hepatocellular carcinoma cells to TRAIL-induced apoptosis via calpain-mediated Bax cleavage. *The international journal of biochemistry & cell biology*. 2017;83:47-55.
  52. Xu Y, Wang Q, Zhang L, Zheng M. 2-Deoxy-D-glucose enhances TRAIL-induced apoptosis in human gastric cancer cells through downregulating JNK-mediated cytoprotective autophagy. *Cancer chemotherapy and pharmacology*. 2018; 81(3):555-64.
  53. Jan R, Chaudhry G-ES. Understanding Apoptosis and Apoptotic Pathways Targeted Cancer Therapeutics. *Adv Pharm Bull*. 2019; 9(2):205-18.
  54. Safa AR. Roles of c-FLIP in Apoptosis, Necroptosis, and Autophagy. *Journal of*

- carcinogenesis & mutagenesis. 2013;Suppl 6.
55. LaCasse EC, Mahoney DJ, Cheung HH, Plenchette S, Baird S, Korneluk RG. IAP-targeted therapies for cancer. *Oncogene*. 2008; 27(48):6252-75.
  56. Bunz F, Hwang PM, Torrance C, Waldman T, Zhang Y, Dillehay L, et al., Disruption of p53 in human cancer cells alters the responses to therapeutic agents. *The Journal of clinical investigation*. 1999;104(3):263-9.
  57. Rustum YM. Thymidylate synthase: a critical target in cancer therapy? *Frontiers in bioscience: a journal and virtual library*. 2004; 9:2467-73.
  58. Deveraux QL, Roy N, Stennicke HR, Van Arsdale T, Zhou Q, Srinivasula SM, et al., IAPs block apoptotic events induced by caspase-8 and cytochrome c by direct inhibition of distinct caspases. *The EMBO journal*. 1998; 17(8):2215-23.
  59. Deveraux QL, Roy N, Stennicke HR, Van Arsdale T, Zhou Q, Srinivasula SM, et al., IAPs block apoptotic events induced by caspase-8 and cytochrome c by direct inhibition of distinct caspases. *The EMBO journal*. 1998; 17(8):2215-23.
  60. Mannhold R, Fulda S, Carosati E. IAP antagonists: promising candidates for cancer therapy. *Drug discovery today*. 2010; 15(5-6):210-9.
  61. Saelens X, Festjens N, Vande Walle L, van Gurp M, van Loo G, Vandenabeele P. Toxic proteins released from mitochondria in cell death. *Oncogene*. 2004; 23(16):2861-74.
  62. Fulda S, Jeremias I, Debatin KM. Cooperation of betulinic acid and TRAIL to induce apoptosis in tumor cells. *Oncogene*. 2004; 23(46):7611-20.
  63. Jan R, Chaudhry GE. Understanding Apoptosis and Apoptotic Pathways Targeted Cancer Therapeutics. *Adv Pharm Bull*. 2019; 9(2):205-18.
  64. Lapenna S, Giordano A. Cell cycle kinases as therapeutic targets for cancer. *Nature reviews Drug discovery*. 2009;8(7):547.
  65. Fathy M, Awale S, Nikaido T. Phosphorylated Akt Protein at Ser473 Enables HeLa Cells to Tolerate Nutrient-Deprived Conditions. *Asian Pacific journal of cancer prevention: APJCP*. 2017;18(12):3255-60.
  66. Strock CJ, Park JI, Nakakura EK, Bova GS, Isaacs JT, Ball DW, et al., Cyclin-dependent kinase 5 activity controls cell motility and metastatic potential of prostate cancer cells. *Cancer research*. 2006;66(15):7509-15.
  67. Khan I, Bahuguna A, Bhardwaj M, Pal Khaket T, Kang SC. Carvacrol nano-emulsion evokes cell cycle arrest, apoptosis induction and autophagy inhibition in doxorubicin resistant-A549 cell line. *Artificial Cells, Nanomedicine, and Biotechnology*. 2018;46(sup1):664-75.
  68. Zhang P, Kawakami H, Liu W, Zeng X, Strebhardt K, Tao K, et al., Targeting CDK1 and MEK/ERK Overcomes Apoptotic Resistance in BRAF-Mutant Human Colorectal Cancer. *Molecular Cancer Research*. 2018;16(3):378.
  69. Humeau J, Bravo-San Pedro JM, Vitale I, Nunez L, Villalobos C, Kroemer G, et al., Calcium signaling and cell cycle: progression or death. *Cell calcium*. 2018; 70:3-15.
  70. Shapovalov G, Skryma R, Prevarskaya N. Calcium channels and prostate cancer. *Recent patents on anti-cancer drug discovery*. 2013;8(1):18-26.
  71. Diaz-Cano SJ. Tumor heterogeneity: mechanisms and bases for a reliable application of molecular marker design. *International journal of molecular sciences*. 2012;13(2):1951-2011.
  72. Su J, Wu S, Wu H, Li L, Guo T. CD44 is functionally crucial for driving lung cancer stem cells metastasis through Wnt/beta-catenin-FoxM1-Twist signaling. *Molecular carcinogenesis*. 2016;55(12):1962-73.
  73. Noto Z, Yoshida T, Okabe M, Koike C, Fathy M, Tsuno H, et al., CD44 and SSEA-4 positive cells in an oral cancer cell line HSC-4 possess cancer stem-like cell characteristics. *Oral oncology*. 2013;49(8):787-95.
  74. Fulda S, Pervaiz S. Apoptosis signaling in cancer stem cells. *The international journal of biochemistry & cell biology*. 2010;42(1):31-8.
  75. Rapp UR, Ceteci F, Schreck R. Oncogene-induced plasticity and cancer stem cells. *Cell cycle*. 2008;7(1):45-51.
  76. Li L, Borodyansky L, Yang Y. Genomic instability en route to and from cancer stem cells. *Cell cycle*. 2009;8(7):1000-2.

77. Dragu DL, Necula LG, Bleotu C, Diaconu CC, Chivu-Economescu M. Therapies targeting cancer stem cells: Current trends and future challenges. *World J Stem Cells*. 2015;7(9):1185-201.
78. Yin S, Xu L, Bandyopadhyay S, Sethi S, Reddy KB. Cisplatin and TRAIL enhance breast cancer stem cell death. *International journal of oncology*. 2011;39(4):891-8.
79. Plasilova M, Zivny J, Jelinek J, Neuwirtova R, Cermak J, Necas E, et al., TRAIL (Apo2L) suppresses growth of primary human leukemia and myelodysplasia progenitors. *Leukemia*. 2002;16(1): 67-73.
80. Unterkircher T, Cristofanon S, Vellanki SH, Nonnenmacher L, Karpel-Massler G, Wirtz CR, et al., Bortezomib primes glioblastoma, including glioblastoma stem cells, for TRAIL by increasing tBid stability and mitochondrial apoptosis. *Clinical cancer research: an official journal of the American Association for Cancer Research*. 2011;17(12):4019-30.
81. Loebinger MR, Sage EK, Davies D, Janes SM. TRAIL-expressing mesenchymal stem cells kill the putative cancer stem cell population. *British journal of cancer*. 2010;103(11):1692-7.
82. Dutta J, Fan Y, Gupta N, Fan G, Gelinas C. Current insights into the regulation of programmed cell death by NF-kappaB. *Oncogene*. 2006;25(51):6800-16.
83. Zhou J, Zhang H, Gu P, Bai J, Margolick JB, Zhang Y. NF-kappaB pathway inhibitors preferentially inhibit breast cancer stem-like cells. *Breast cancer research and treatment*. 2008;111(3):419-27.
84. Guzman ML, Swiderski CF, Howard DS, Grimes BA, Rossi RM, Szilvassy SJ, et al., Preferential induction of apoptosis for primary human leukemic stem cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2002;99(25):16220-5.
85. Fathy M, Nikaido T. In vivo modulation of iNOS pathway in hepatocellular carcinoma by *Nigella sativa*. *Environmental health and preventive medicine*. 2013;18(5):377-85.
86. Fathy M, Nikaido T. In vivo attenuation of angiogenesis in hepatocellular carcinoma by *Nigella sativa*. *Turkish journal of medical sciences*. 2018;48(1):178-86.
87. Fathy M, Khalifa E, Fawzy MA. Modulation of inducible nitric oxide synthase pathway by eugenol and telmisartan in carbon tetrachloride-induced liver injury in rats. *Life sciences*. 2019;216:207-14.
88. Kronski E, Fiori ME, Barbieri O, Astigiano S, Mirisola V, Killian PH, et al., miR181b is induced by the chemopreventive polyphenol curcumin and inhibits breast cancer metastasis via down-regulation of the inflammatory cytokines CXCL1 and -2. *Molecular oncology*. 2014;8(3):581-95.
89. Charpentier MS, Whipple RA, Vitolo MI, Boggs AE, Slovic J, Thompson KN, et al., Curcumin targets breast cancer stem-like cells with microtentacles that persist in mammospheres and promote reattachment. *Cancer research*. 2014;74(4):1250-60.
90. Park HY, Toume K, Arai MA, Sadhu SK, Ahmed F, Ishibashi M. Calotropin: a cardenolide from *calotropis gigantea* that inhibits Wnt signaling by increasing casein kinase 1alpha in colon cancer cells. *Chembiochem: a European journal of chemical biology*. 2014;15(6):872-8.
91. Singh SV, Singh K. Cancer chemoprevention with dietary isothiocyanates mature for clinical translational research. *Carcinogenesis*. 2012;33(10):1833-42.
92. Rausch V, Liu L, Kallifatidis G, Baumann B, Mattern J, Gladkich J, et al., Synergistic activity of sorafenib and sulforaphane abolishes pancreatic cancer stem cell characteristics. *Cancer research*. 2010; 70(12):5004-13.
93. Li Y, Zhang T, Korkaya H, Liu S, Lee HF, Newman B, et al., Sulforaphane, a dietary component of broccoli/broccoli sprouts, inhibits breast cancer stem cells. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2010;16(9):2580-90.
94. Lee HA, Park S, Kim Y. Effect of beta-carotene on cancer cell stemness and differentiation in SK-N-BE(2)C neuroblastoma cells. *Oncology reports*. 2013;30(4): 1869-77.
95. Li J-f, Niu Y-y, Xing Y-l, Liu F. A novel bispecific c-MET/CTLA-4 antibody targeting lung cancer stem cell-like cells with therapeutic potential in human non-small-cell lung cancer. *Bioscience Reports*. 2019;39(5).

96. Agwa ES, Ma PC. Targeting the MET receptor tyrosine kinase in non-small cell lung cancer: emerging role of tivantinib. *Cancer management and research*. 2014; 6:397-404.
97. Liu Y, Wang T, Yan J, Jiagbogu N, Heideman DA, Canfield AE, et al., HGF/c-Met signalling promotes Notch3 activation and human vascular smooth muscle cell osteogenic differentiation in vitro. *Atherosclerosis*. 2011;219(2):440-7.
98. Sharma N, Nanta R, Sharma J, Gunewardena S, Singh KP, Shankar S, et al., PI3K/AKT/mTOR and sonic hedgehog pathways cooperate together to inhibit human pancreatic cancer stem cell characteristics and tumor growth. *Oncotarget*. 2015;6(31):32039-60.
99. Nakai E, Park K, Yawata T, Chihara T, Kumazawa A, Nakabayashi H, et al., Enhanced MDR1 expression and chemoresistance of cancer stem cells derived from glioblastoma. *Cancer investigation*. 2009;27(9):901-8.
100. Safa AR. Resistance to Cell Death and Its Modulation in Cancer Stem Cells. *Critical reviews in oncogenesis*. 2016;21(3-4):203-19.
101. Shang C, Guo Y, Hong Y, Liu Y-h, Xue Y-x. MiR-21 up-regulation mediates glioblastoma cancer stem cells apoptosis and proliferation by targeting FASLG. *Molecular Biology Reports*. 2015;42(3): 721-7.
102. Liang-Hu Q. RNomics: the new frontier in the post-genomic era. *Science in China Series C, Life sciences*. 2009;52(3):193-4.
103. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell*. 2009;136(2):215-33.
104. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell*. 2004;116(2):281-97.
105. Indran IR, Tufo G, Pervaiz S, Brenner C. Recent advances in apoptosis, mitochondria and drug resistance in cancer cells. *Biochimica et biophysica acta*. 2011;1807(6):735-45.
106. Iorio MV, Croce CM. MicroRNAs in cancer: small molecules with a huge impact. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2009;27(34):5848-56.
107. Zhou K, Liu M, Cao Y. New insight into microRNA functions in cancer: oncogene-microRNA-tumor suppressor gene network. *Frontiers in molecular biosciences*. 2017;4:46.
108. Gandellini P, Profumo V, Folini M, Zaffaroni N. MicroRNAs as new therapeutic targets and tools in cancer. *Expert opinion on therapeutic targets*. 2011;15(3): 265-79.
109. Kota J, Chivukula RR, O'Donnell KA, Wentzel EA, Montgomery CL, Hwang HW, et al., Therapeutic microRNA delivery suppresses tumorigenesis in a murine liver cancer model. *Cell*. 2009;137(6): 1005-17.
110. Bonci D, Coppola V, Musumeci M, Addario A, Giuffrida R, Memeo L, et al., The miR-15a-miR-16-1 cluster controls prostate cancer by targeting multiple oncogenic activities. *Nature medicine*. 2008;14(11):1271-7.
111. Xiong Y, Fang JH, Yun JP, Yang J, Zhang Y, Jia WH, et al., Effects of microRNA-29 on apoptosis, tumorigenicity, and prognosis of hepatocellular carcinoma. *Hepatology*. 2010;51(3):836-45.
112. Qin W, Shi Y, Zhao B, Yao C, Jin L, Ma J, et al., miR-24 regulates apoptosis by targeting the open reading frame (ORF) region of FAF1 in cancer cells. *PloS one*. 2010;5(2):e9429.
113. De Guire V, Caron M, Scott N, Menard C, Gaumont-Leclerc MF, Chartrand P, et al., Designing small multiple-target artificial RNAs. *Nucleic acids research*. 2010;38(13):e140.
114. Singh R, Saini N. Downregulation of BCL2 by miRNAs augments drug-induced apoptosis--a combined computational and experimental approach. *Journal of cell science*. 2012;125(Pt 6):1568-78.
115. Cimmino A, Calin GA, Fabbri M, Iorio MV, Ferracin M, Shimizu M, et al., miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proceedings of the National Academy of Sciences of the United States of America*. 2005;102(39):13944-9.
116. Johnson CD, Esquela-Kerscher A, Stefani G, Byrom M, Kelnar K, Ovcharenko D, et al., The let-7 microRNA represses cell proliferation pathways in human cells. *Cancer research*. 2007;67(16):7713-22.

117. Bottoni A, Piccin D, Tagliati F, Luchin A, Zatelli MC, degli Uberti EC. miR-15a and miR-16-1 down-regulation in pituitary

adenomas. Journal of cellular physiology. 2005;204(1):280-5.